AMENDMENTS TO THE SPECIFICATION

Docket No.: 13987-00022-US

Please delete the sequence listing from the English translation of the international application and replace it with the sequence listing submitted on compact disc enclosed herewith.

In the specification at page 1, after the title and before line 4, please insert the following new paragraphs:

RELATED APPLICATIONS

This application is a national stage application (under 35 U.S.C. 371) of PCT/EP2005/002734 filed March 15, 2005, which claims benefit of European application 04006358.8 filed March 17, 2004.

SUBMISSION ON COMPACT DISC

The contents of the following submission on compact discs are incorporated herein by reference in its entirety: two copies of the Sequence Listing (COPY 1 and COPY 2) and a computer readable form copy of the Sequence Listing (CRF COPY), all on compact disc, each containing: file name: Final Sequence List-13987-00022-US, date recorded: September 14, 2006, size: 102 KB.

In the specification at page 9 line 34, please replace the paragraph starting with "The Damino acid" with the following amended paragraph:

The D-amino acid oxidase expressed from the DNA-construct of the invention has preferably metabolising activity against at least one D-amino acid and comprises a sequences motive having the following consensus sequence (SEQ ID NO: 17):

In the specification at page 33 line 14, please replace the paragraph starting with "In DAAO" with the following amended paragraph:

In DAAO, a conserved histidine has been shown (Miyano M et al. (1991) J Biochem 109:171-177) to be important for the enzyme's catalytic activity. In a preferred embodiment of the invention a DAAO is referring to a protein comprising the following consensus motive (SEQ ID NO: 17):

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In the specification at page 49 line 12, please replace Table 4 with the following amended Table 4:

Recombi-	Organism	Recombination Sites	SEQ ID
nase	of origin		NO:
CRE	Bacteriophage	5'-AACTCTCATCGCTTCGGATAACTTCCTGTTATCCGAAA	<u>18</u>
	P1	CATATCACTCACTTTGGTGATTTCACCGTAACT-	
		GTCTATGATTAATG-3'	
FLP	Saccharomyces	5'-GAAGTTCCTATTCCGAAGTTCCTATTCTCTAGAA AG-	<u>19</u>
	cerevisiae	TATAGGAACTTC-3'	
R	pSR1	5'-CGAGATCATATCACTGTGGACGTTGATGAAAGAATAC	20
	Plasmids	GTTATTCTTTCATCAAATCGT	

In the specification starting at page 56 through to page 59, please replace Table 5 with the following amended Table 5:

DSBI	Organism	Recognition sequence	SEQ ID
Enzyme	of origin		<u>NO :</u>
P-	Drosophila	5'-CTAGATGAAATAACATAAGGTGG	<u>21</u>
Element	·		
Trans-			
posase			
I-Anil	Aspergillus nidu-		22
	lans	3'-AACTCCTCCAAAGAGACATTTATTNNNNNNNNNNNNNNNN	
I-Ddil	Dictyostelium	5'-TTTTTTGGTCATCCAGAAGTATAT	<u>23</u>
	discoideumAX3	3'-AAAAAACCAG^TAGGTCTTCATATA	
I-Cvul	Chlorella vulgaris	5'-CTGGGTTCAAAACGTCGTGA^GACAGTTTGG	24
		3'-GACCCAAGTTTTGCAG^CACTCTGTCAAACC	
I-Csml	Chlamydomonas	5'-GTACTAGCATGGGGTCAAATGTCTTTCTGG	<u>25</u>
	smithii		

DSBI	Organism	Recognition sequence	SEQ ID
Enzyme	of origin		<u>NO :</u>
I-Cmoel	Chlamydomonas	5'-TCGTAGCAGCT^CACGGTT	<u>26</u>
}	moewusii	3'-AGCATCG^TCGAGTGCCAA	
I-Crel	Chlamydomonas	5'-CTGGGTTCAAAACGTCGTGA^GACAGTTTGG	<u>27</u>
	reinhardtii	3'-GACCCAAGTTTTGCAG^CACTCTGTCAAACC	
I-Chul	Chlamydomonas	5'-GAAGGTTTGGCACCTCG^ATGTCGGCTCATC	28
	humicola	3'-CTTCCAAACCGTG^GAGCTACAGCCGAGTAG	
I-Cpal	Chlamydomonas	5'-CGATCCTAAGGTAGCGAA^ATTCA	<u>29</u>
-	pallidostigmatica	3'-GCTAGGATTCCATC^GCTTTAAGT	
I-Cpall	Chlamydomonas	5'-CCCGGCTAACTC^TGTGCCAG	<u>30</u>
-	pallidostigmatica	3'-GGGCCGAT^TGAGACACGGTC	
I-Ceul	Chlamydomonas	5'-CGTAACTATAACGGTCCTAA^GGTAGCGAA	31
	eugametos	3'-GCATTGATATTGCCAG^GATTCCATCGCTT	
I-Dmol	Desulfuro-	5'-ATGCCTTGCCGGGTAA^GTTCCGGCGCGCAT	<u>32</u>
	coccus mobilis	3'-TACGGAACGGCC^CATTCAAGGCCGCGCGTA	
I-Scel	Saccharomyces	5'-AGTTACGCTAGGGATAA^CAGGGTAATATAG	<u>33</u>
	cerevisiae	3'-TCAATGCGATCCC^TATTGTCCCATTATATC	
		5'-TAGGGATAA^CAGGGTAAT	34
		3'-ATCCC^TATTGTCCCATTA ("Core"-Sequence)	
I-Scell	S.cerevisiae	5'-TTTTGATTCTTTGGTCACCC^TGAAGTATA	<u>35</u>
		3'-AAAACTAAGAAACCAG^TGGGACTTCATAT	
I-Scelll	S.cerevisiae	5'-ATTGGAGGTTTTGGTAAC^TATTTATTACC	<u>36</u>
10 01		3'-TAACCTCCAAAACC^ATTGATAAATAATGG	27
I-ScelV	S.cerevisiae	5'-TCTTTTCTCTTGATTA^GCCCTAATCTACG 3'-AGAAAAGAGAAC^TAATCGGGATTAGATGC	<u>37</u>
I Cool/	Correvision	5'-AATAATTTTCT^TCTTAGTAATGCC	38
I-SceV	S.cerevisiae	3'-TTATTAAAAGAAGAATCATTA^CGG	30
I-SceVI	S.cerevisiae	5'-GTTATTTAATG^TTTTAGTAGTTGG	39
1-006 41	O.CCI CVISIAC	3'-CAATAAATTACAAAATCATCA^ACC	
I-SceVII	S.cerevisiae	5'-TGTCACATTGAGGTGCACTAGTTATTAC	40
PI-Scel	S.cerevisiae	5'-ATCTATGTCGGGTGC^GGAGAAAGAGGTAAT	41
		3'-TAGATACAGCC^CACGCCTCTTTCTCCATTA	
F-Scel	S.cerevisiae	5'-GATGCTGTAGGC^ATAGGCTTGGTT	42
		3'-CTACGACA^TCCGTATCCGAACCAA	
F-Scell	S.cerevisiae	5'-CTTTCCGCAACA^GTAAAATT	43
		3'-GAAAGGCG^TTGTCATTTTAA	
I-Hmul	Bacillus subtilis	5'-AGTAATGAGCCTAACGCTCAGCAA	44
	bacteriophage	3'-TCATTACTCGGATTGC^GAGTCGTT	
	SPO1		

DSBI	Organism	Recognition sequence	SEQ ID
Enzyme	of origin		NO:
I-Hmull	Bacillus subtilis bacteriophage SP82	5'-AGTAATGAGCCTAACGCTCAACAANNNNNNNNNNNNNNNN	<u>45</u>
I-Llal	Lactococcus lactis	5'-CACATCCATAAC^CATATCATTTTT 3'-GTGTAGGTATTGGTATAGTAA^AAA	46
I-Msol	Monomastix species	5'-CTGGGTTCAAAACGTCGTGA^GACAGTTTGG 3'-GACCCAAGTTTTGCAG^CACTCTGTCAAACC	47
I-Nanl	Naegleria andersoni	5'-AAGTCTGGTGCCA^GCACCCGC 3'-TTCAGACC^ACGGTCGTGGGCG	48
I-Nitl	Naegleria italica	5'-AAGTCTGGTGCCA^GCACCCGC 3'-TTCAGACC^ACGGTCGTGGGCG	<u>49</u>
I-Njal	Naegleria jamiesoni	5'-AAGTCTGGTGCCA^GCACCCGC 3'-TTCAGACC^ACGGTCGTGGGCG	<u>50</u>
I-Pakl	Pseudendoclonium akinetum	5'-CTGGGTTCAAAACGTCGTGA^GACAGTTTGG 3'-GACCCAAGTTTTGCAG^CACTCTGTCAAACC	<u>51</u>
I-Porl	Pyrobaculum organotrophum	5'-GCGAGCCCGTAAGGGT^GTGTACGGG 3'-CGCTCGGGCATT^CCCACACATGCCC	<u>52</u>
I-Ppol	Physarum polycephalum	5'-TAACTATGACTCTCTTAA^GGTAGCCAAAT 3'-ATTGATACTGAGAG^AATTCCATCGGTTTA	<u>53</u>
I-Scal	Saccharomyces capensis	5'-TGTCACATTGAGGTGCACT^AGTTATTAC 3'-ACAGTGTAACTCCAC^GTGATCAATAATG	<u>54</u>
I- Ssp6803I	Synechocystis species	5'-GTCGGGCT^CATAACCCGAA 3'-CAGCCCGAGTA^TTGGGCTT	<u>55</u>
PI-Pful	Pyrococcus furiosus Vc1	5'-GAAGATGGGAGGGGACCGGACTCAACTT 3'-CTTCTACCCTCC^TCCCTGGCCTGAGTTGAA	<u>56</u>
PI-Pfull	Pyrococcus furiosus Vc1	5'-ACGAATCCATGTGGAGA^AGAGCCTCTATA 3'-TGCTTAGGTACAC^CTCTTCTCGGAGATAT	<u>57</u>
PI-Pkol	Pyrococcus kodakaraensis KOD1	5'-GATTTTAGAT^CCCTGTACC 3'-CTAAAA^TCTAGGGACATGG	<u>58</u>
PI-Pkoll	Pyrococcus kodakaraensis KOD1	5'-CAGTACTACG^GTTAC 3'-GTCATG^ATGCCAATG	<u>59</u>
PI-Pspl	Pyrococcus sp.	5'-AAAATCCTGGCAAACAGCTATTAT^GGGTAT 3'-TTTTAGGACCGTTTGTCGAT^AATACCCATA	<u>60</u>

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DSBI	Organism	Recognition sequence	SEQ ID
Enzyme	of origin		NO:
PI-Tful	Thermococcus	5'-TAGATTTTAGGT^CGCTATATCCTTCC	<u>61</u>
	fumicolans ST557	3'-ATCTAAAA^TCCAGCGATATAGGAAGG	
PI-Tfull	Thermococcus	5'-TAYGCNGAYACN^GACGGYTTYT	<u>62</u>
	fumicolans ST557	3'-ATRCGNCT^RTGNCTGCCRAARA	
PI-Thyl	Thermococcus	5'-TAYGCNGAYACN^GACGGYTTYT	<u>62</u>
	hydrothermalis	3'-ATRCGNCT^RTGNCTGCCRAARA	
PI-TIII	Thermococcus	5'-TAYGCNGAYACNGACGG^YTTYT	<u>62</u>
	litoralis	3'-ATRCGNCTRTGNC^TGCCRAARA	
PI-TIII	Thermococcus	5'-AAATTGCTTGCAAACAGCTATTACGGCTAT	<u>63</u>
	litoralis		
I-Tevl	Bacteriophage T4	5'-AGTGGTATCAAC^GCTCAGTAGATG	<u>64</u>
		3'-TCACCATAGT^TGCGAGTCATCTAC	
I-TevII	Bacteriophage T4	5'-GCTTATGAGTATGAAGTGAACACGT^TATTC	<u>65</u>
		3'-CGAATACTCATTCACTTGTG^CAATAAG	
F-TevI	Bacteriophage T4	5'-GAAACACAAGA^AATGTTTAGTAAANNNNNNNNNNNNNNNNN	<u>66</u>
		3'-CTTTGTGTTCTTTACAAATCATTTNNNNNNNNNNNNNNNN	
F-TevII	Bacteriophage T4	5'-TTTAATCCTCGCTTC^AGATATGGCAACTG	<u>67</u>
		3'-AAATTAGGAGCGA^AGTCTATACCGTTGAC	
H-Drel	E. coli pl-Drel	5'-CAAAACGTCGTAA^GTTCCGGCGCG	<u>68</u>
1.0		3'-GTTTTGCAG^CATTCAAGGCCGCGC	60
I-Basl	Bacillus	5' AGTAATGAGCCTAACGCTCAGCAA	<u>69</u>
	thuringiensis	3'- TCATTACGAGTCGAACTCGGATTG	<u>70</u>
	phage Bastille		
I-Bmol	Bacillus	5'-GAGTAAGAGCCCG^TAGTAATGACATGGC	<u>71</u>
	mojavensis s87-18	3'-CTCATTCTCG^GGCATCATTACTGTACCG	
I-Pogl	Pyrobaculum	5'-CTTCAGTAT^GCCCCGAAAC	72
	oguniense	3'-GAAGT^CATACGGGGCTTTG	
I-Twol	Staphylococcus	5'-TCTTGCACCTACACAATCCA	<u>73</u>
	aureus phage	3'-AGAACGTGGATGTTAGGT	
	Twort		į
PI-Mgal	Mycobacterium	5'-CGTAGCTGCCCAGTATGAGTCA	74
J	gastri	3'-GCATCGACGGGTCATACTCAGT	
PI-Pabl	Pyrococcus abyssi	5'-GGGGCAGCCAGTGGTCCCGTT	<u>75</u>
		3'-CCCCGTCGGTCACCAGGGCAA	
PI-Pabli	Pyrococcus abyssi	5'-ACCCCTGTGGAGAGGAGCCCCTC	<u>76</u>
		3'-TGGGGACACCTCTCCTCGGGGAG	

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In the specification at page 89 line 1, please replace the paragraph starting with "Fig.: 12" with the following amended paragraph:

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Fig.: 12 Alignment of the catalytic site of various D-amino acid oxidases

Multiple alignment of the catalytic site of various D-amino acid oxidases allows for
determination of a characteristic sequence motif [LIVM]-[LIVM]-H*-[NHA]-Y-G-x[GSA]-[GSA]-x-G-x₅-G-x-A (SEQ ID NO: 17), which allows for easy identification of
additional D-amino acid oxidases suitable to be employed within the method and DNAconstructs of the invention.

In the specification at page 90 line 32, please replace the paragraph starting with "The yeast" with the following amended paragraph:

The yeast R. gracilis was grown in liquid culture containing 30 mM D-alanine to induce dao1, the gene encoding DAAO. Total RNA was isolated from the yeast and used for cDNA synthesis. The PCR primers

- 5'-ATTAGATCTTACTACTCGAAGGACGCCATG-3' (SEQ ID NO: 77) and
- 5'-ATTAGATCTACAGCCACAATTCCCGCCCTA-3' (SEQ ID NO: 78)